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## $\alpha$ -Lipoic acid protects against $\gamma$ -rays-induced cardiac injury in rats

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$\alpha$ -LIPOIC acid is found in some mammalian and plant tissues. It has antioxidant and anti-inflammatory effects. The objective of this study was to test the hypothesis that oral administration of  $\alpha$ -lipoic acid causes protection against cardiac injury in  $\gamma$ -irradiated rats.

A group of rats was exposed to a single session of 6 Gy  $\gamma$ -rays and compared with three other groups: a control group (received vehicle only), an  $\alpha$ -lipoic acid-treated group (received 100 mg/kg body weight orally for 10 days), and a combined treatment group (received 6 Gy  $\gamma$ -radiation along with  $\alpha$ -lipoic acid for 10 days).

The effect of radiation was indicated by an increase in the activity of lactate dehydrogenase (LDH), creatin Kinase MB (CK-MB), myeloperoxidase (MPO), xanthine oxidase (XO), malondialdehydes (MDA), protein carbonyl (PC), and cytokine IL-6 (IL-6) and a significant decrease in the activity of superoxide dismutase (SOD), glutathione peroxidase (GPx), catalase (CAT), ferric reducing ability (FRA) and reduced glutathione (GSH) concentrations.

$\alpha$ -Lipoic acid supplementation to  $\gamma$ -rays group ameliorates all parameter alterations occurred in blood and rat cardiac tissues, suggesting the existence of a compensatory increase of antioxidant defences.

It could be postulated that  $\alpha$ -lipoic, as a functional dietary supplement, could exert a modulator role in the radiation-induced cardiac injury through its antioxidant and anti-inflammatory properties.

**Keywords:**  $\alpha$ -Lipoic acid,  $\gamma$ -rays, myocardium, rats

### Introduction

Acute exposure to  $\gamma$ -radiation (6 Gy) results in cardiac injury in rats and triggers cardiac inflammation (Karam et al., 2024). Gamma-ray-induced cardiac disease is a common complication of thoracic radiotherapy (Liu et al., 2025). In rats, ionising radiation is associated with oxidative stress and impaired immune function (Aziz et

al., 2024; Hussien and Rashed, 2023). Moreover, gamma rays enhance lipid peroxidation in rat cells (Moselhy et al., 2025). Oxidative stress, reactive oxygen species (ROS), and hypoxia contribute to cardiac dysfunction and acute myocardial ischaemia injury (Jiang et al., 2025).

$\alpha$ -Lipoic acid is a naturally occurring molecule found in certain mammalian and plant tissues.

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It mitigates pro-oxidant and pro-inflammatory activities and modulates both nitrogen and ROS species (Pingali *et al.*, 2024). Supplementation with  $\alpha$ -lipoic acid may offer protection against immune dysfunction and cardiac injury in mice (Wang *et al.*, 2023). Additionally, it functions as a free radical scavenger and metal-chelating compound (Pang *et al.*, 2024).  $\alpha$ -Lipoic acid also alleviates toxins associated with oxidative stress-related diseases (Zhao *et al.*, 2025).

$\alpha$ -Lipoic acid has proven effective in reducing the extent of myocardial injury (Wang *et al.*, 2025<sup>b</sup>). In both rats and mice, it demonstrates a safe and protective role against myocardial ischaemia-reperfusion injury (Oskuye *et al.*, 2024; Wang *et al.*, 2025<sup>b</sup>).

$\alpha$ -Lipoic acid provides protection against myocardial ischaemia-reperfusion injury by directly interacting with lipid peroxidation products (Sztolsztener *et al.*, 2022), detoxifying aldehydes generated from oxidised lipids (Khan *et al.*, 2022). Furthermore, its cardioprotective efficacy following myocardial infarction in humans has been demonstrated (Jermendy *et al.*, 2023). In animal studies,  $\alpha$ -lipoic acid has also shown effective therapeutic potential in treating myocardial perfusion injury in rats and mice (Xie *et al.*, 2023; Wang *et al.*, 2025<sup>b</sup>). In addition,  $\alpha$ -lipoic acid prevents  $\gamma$ -ray-induced testicular injury by restoring normal testicular function, ultimately leading to the recovery of spermatogenesis in rats (Said *et al.*, 2020). Moreover, it protects against  $\gamma$ -ray-induced brain injury in rats by reactivating antioxidant and anti-inflammatory mechanisms (Xu *et al.*, 2023).

It has been postulated that  $\alpha$ -lipoic acid administration may offer protection against streptozotocin/high-fat diet-induced diabetic nephropathy (Helmy *et al.*, 2024). Furthermore,  $\alpha$ -lipoic acid exhibits a protective effect against ultraviolet-A/B-induced skin damage (Tülüce *et al.*, 2024). It may also provide a novel preventive approach due to its potent antioxidant and anti-inflammatory properties, alongside its favourable safety profile in the lungs of irradiated rats (Sayedpour *et al.*, 2024). Additionally,  $\alpha$ -lipoic acid mitigates radiotherapy-induced liver tissue damage (Gezer *et al.*, 2023), and its pretreatment has been shown to alleviate ionising radiation-induced oxidative stress and apoptosis in rat brain tissue (Xu *et al.*, 2023).

The purpose of this article is studying the

efficacy of  $\alpha$ -lipoic acid on cardiac injury induced in whole body gamma-irradiated rats.

## Material and Methods

### Experimental animals

Thirty-two male Wister Rats 9-10 weeks old, weighing  $335 \pm 11$ g were used. They were bought from the Holding Company for Biological Products and Vaccines (VACSERA, Helwan, Cairo, Egypt). The experimental animals were kept in suitable cages and controlled environmental situation, constant temperature (24-28 °C), constant light cycle (light from 6.00<sub>AM</sub> to 6.00<sub>PM</sub>), and humidity (50-70%). Rats were supplied normal balanced rat-pellets as well as water *ad-libitum*. The animals were kept and used in agreement with the animal ethics. All experiments were conformed to the procedure and guides for the care and use of laboratory animals published earlier, National Institutes of Health, No. 85-23, revised 1996.

### Radiation technique

Irradiation was performed with a Canadian gamma cell-40, ( $^{137}\text{Cs}$ ) at the NCRRT, Cairo, Egypt. Animals were exposed to one session of  $\gamma$ -rays at a dose rate 0.37 Gy/ minute. The selected  $\gamma$ -rays dose (6Gy) was enough to induce distinct biochemical alteration and pathological lesion (Mekkawy *et al.*, 2024; Tawfik *et al.*, 2019).

### Animal groups and treatment schedules

Rats were divided into four groups, each group 8rats. Control group: Rats were received 1ml bi-distilled water daily (as a vehicle) / for ten following days via stomach-tubes.  $\alpha$ -lipoic acid group: Rats were received  $\alpha$ -lipoic acid (Sigma-Aldrich, USA) suspended in 1ml of the vehicle (100mg/ kg/ day for 10days) via stomach-tubes, according to Abdel-Aziz *et al.* (2021) and Andreeva *et al.* (2020).  $\gamma$ -rays group: Rats were exposed to acute whole body  $\gamma$ -rays (6Gy-one single dose) then after half an hour, rats received the vehicle daily/ for ten days via stomach-tubes.  $\gamma$ -rays+  $\alpha$ -lipoic acid group: Rats were exposed to  $\gamma$ -rays (6Gy) then were received via stomach-tubes  $\alpha$ -lipoic acid (100mg) half an hour post-irradiation and continued for ten days.

Rats in all groups were killed 24hours after the end of the experiment period. The blood plasma and sera were prepared from collected whole blood samples from rat hearts. The hearts were quickly removed. The heart muscles were kept on ice; they were then trimmed from fat and connective tissue and instantly frozen at -80 °C.

*Biochemical measures in blood and cardiac tissue*

Lactate dehydrogenase (LDH) and creatin Kinase MB (CK-MB) activities were estimated in serum as an induces for cardiac cells damage induced by gamma-radiation. LDH (E.C.1.1.1.27) and CK-MB (Catalog Number: 239 002) activities were determined at absorbances of 340 nm using kinetic methods distributed by Spectrum-diagnosis, Cairo, Egypt. The cardiac tissue myeloperoxidase (MPO); A044-1-1 was estimated according to the patent kit of Jianchen, Nanjing, China. The xanthine oxidase (XO) was estimated according to Bergmeyer et al. (1983). The malondialdehydes (MDA) assessed as described by Buege and Aust (1987). The quantification of protein carbonyl (PC) was assessed according to Levine et al. (1990) technique. The plasma ferric reducing ability (FRA) level was assessed as Bustamante et al. (1998) described. The serum level of cytokine IL-6 (IL-6) was estimated by using kit (Catalog Number: CSB-E04640r) obtained from CUSABIO Technology Biotech Company, China. The cardiac superoxide dismutase (SOD), glutathione peroxidase (GPx) and Catalase (CAT) activities were measured according to Lawrence and Burk (2012), Kakkar et al. (1984), Titov and Osipoc (2017), and reduced glutathione (GSH) content was estimated according to Cigala et al. (2012). The amount of total heart protein was performed according Lowry et al. (1951) method.

*Statistical analysis*

The obtained data were expressed as mean  $\pm$  standard error (SE). Statistical analysis was performed using one-way analysis of variance (ANOVA), followed by Duncan's post hoc test to determine significant differences among the experimental groups. The normality of data

distribution was assessed using the Shapiro-Wilk test. Statistical analyses were conducted using SPSS software. A *p*-value of less than 0.05 was considered statistically significant.

**Results**

All experimental investigations and statistical tests were carried out blindly. All Groups were given code numbers. No one knows the group of rats during analysis.

The activities of LDH and CK-MB were significantly elevated in the  $\gamma$ -rays group, reaching 1.4- and 2.0-fold increases, respectively, compared to the control, and 1.6- and 2.0-fold increases, respectively, compared to the  $\alpha$ -lipoic acid-treated group. However, these enzyme activities were significantly reduced in the  $\gamma$ -rays+  $\alpha$ -lipoic acid group compared to the  $\gamma$ -rays group and were nearly restored to control levels (Table 1).

MPO and XO activities were significantly increased in the  $\gamma$ -rays group by 4.3- and 2.4-fold, respectively, compared to the control, and by 4.2- and 2.4-fold, respectively, compared to the  $\alpha$ -lipoic acid group.  $\alpha$ -Lipoic acid supplementation reduced both MPO and XO activities in the  $\gamma$ -rays+  $\alpha$ -lipoic acid group, restoring them to near-control levels (Table 1).

In the  $\gamma$ -rays group, MDA, PC, and IL-6 concentrations significantly increased by approximately 2.4-, 2.0-, and 1.7-fold, respectively, when compared to both the control and  $\alpha$ -lipoic acid groups. These levels significantly declined in the  $\gamma$ -rays+  $\alpha$ -lipoic acid group, approaching values similar to the control (Table 2).

**TABLE 1. Serum lactate dehydrogenase (LDH) and creatin kinase MB (CK-MB) and cardiac myeloperoxidase (MPO) and xanthin oxidase (XO) levels after  $\alpha$ -lipoic acid/ or  $\gamma$ -rays exposure in different rat groups.**

Groups	LDH	CK-MB	MPO	XO
	U/ ml	U/ ml	U/ 100 mg tissue	U/ mg protein
Control	271.21 $\pm$ 8.332	148.21 $\pm$ 4.114	5.12 $\pm$ 0.128	0.66 $\pm$ 0.014
$\alpha$ -lipoic acid	266.82 $\pm$ 8.123	146.32 $\pm$ 3.521	5.23 $\pm$ 0.132	0.67 $\pm$ 0.022
$\gamma$ -rays	438.22 $\pm$ 14.651 <sup>a,b</sup>	294.24 $\pm$ 10.444 <sup>a,b</sup>	22.14 $\pm$ 0.705 <sup>a,b</sup>	1.58 $\pm$ .036 <sup>a,b</sup>
$\gamma$ -rays+ $\alpha$ -lipoic acid	308.13 $\pm$ 8.253 <sup>c</sup>	162.12 $\pm$ 4.925 <sup>c</sup>	8.32 $\pm$ 0.205 <sup>c</sup>	0.81 $\pm$ 0.021 <sup>c</sup>

<sup>a</sup> significant difference when comparing with control group.

<sup>b</sup> significant difference when comparing with  $\alpha$ -lipoic acid-treated group.

<sup>c</sup> significant difference when comparing with  $\gamma$ -rays (6Gy) exposure group.

In the  $\alpha$ -lipoic acid group, FRA levels significantly increased compared to the control (1.6-fold). In contrast, the  $\gamma$ -rays group showed a significant reduction in FRA compared to both the control ( $-40.7\%$ ) and  $\alpha$ -lipoic acid-treated ( $-61.9\%$ ) groups. In the  $\gamma$ -rays +  $\alpha$ -lipoic acid group, FRA levels significantly increased relative to both the control (1.5-fold) and  $\gamma$ -rays groups (2.5-fold), with no significant difference observed between the  $\alpha$ -lipoic acid-treated and  $\gamma$ -rays +  $\alpha$ -lipoic acid groups (Table 2).

In the  $\gamma$ -rays group, all oxidative stress markers; SOD, GPx, and CAT activities, along with GSH content were significantly decreased compared to the control group (by  $-35.0\%$ ,  $-54.1\%$ ,  $-62.2\%$ , and  $-55.2\%$ , respectively) and the  $\alpha$ -lipoic acid-treated group (by  $-32.6\%$ ,  $-44.3\%$ ,  $-62.7\%$ , and  $-54.6\%$ , respectively).

In the  $\gamma$ -rays+  $\alpha$ -lipoic acid group, SOD and GPx activities were significantly elevated

compared to the  $\gamma$ -rays group (by 1.4-fold, respectively), and their levels were nearly restored to those of the control. However, although CAT activity and GSH content were also significantly increased compared to the  $\gamma$ -rays group, they remained significantly lower than those in the control group (by  $-23.0\%$  and  $-13.9\%$ , respectively) and the  $\alpha$ -lipoic acid-treated group (by  $-24.0\%$  and  $-12.9\%$ , respectively).

### Discussion

In recent years, gamma radiation has emerged as a contributing factor to cardiac injury during radiotherapy protocols (Liu et al., 2025). Concurrently, potent antioxidants such as  $\alpha$ -lipoic acid have garnered attention for their cardioprotective potential in radiation-induced injuries (Adhab et al., 2025; Wang et al., 2025<sup>b</sup>). The present study aimed to evaluate the protective efficacy of  $\alpha$ -lipoic acid against  $\gamma$ -radiation-induced cardiac damage in rats. Owing to its

**TABLE 2. Cardiac malondialdehydes (MDA) and protein carbonyl (PC), plasma ferric reducing ability (FRA) and serum level of cytokine IL-6 (IL-6) levels after  $\alpha$ -lipoic acid/ or  $\gamma$ -rays exposure in different rat groups.**

Groups	MDA	PC	FRA	IL-6
	$\mu\text{mol/ mg tissue}$	$\text{nmol/ mg tissue}$	$\text{mM/ L}$	$\text{Pg/ ml}$
Control	$2.4 \pm 0.08$	$0.54 \pm 0.018$	$0.86 \pm 0.021$	$44.12 \pm 1.103$
$\alpha$ -lipoic acid	$2.3 \pm 0.08$	$0.54 \pm 0.019$	$1.34 \pm 0.043^a$	$43.59 \pm 1.092$
$\gamma$ -rays	$5.8 \pm 0.28^{a,b}$	$1.09 \pm 0.045^{a,b}$	$0.51 \pm 0.013^{a,b}$	$73.57 \pm 2.355^{a,b}$
$\alpha$ -lipoic acid+ $\gamma$ -rays	$2.6 \pm 0.12^c$	$0.64 \pm 0.021^c$	$1.30 \pm 0.034^{a,c}$	$52.12 \pm 1.274^c$

<sup>a</sup> significant difference when comparing with control group.

<sup>b</sup> significant difference when comparing with  $\alpha$ -lipoic acid-treated group.

<sup>c</sup> significant difference when comparing with  $\gamma$ -rays (6Gy) exposure group.

**TABLE 3. Cardiac superoxide dismutase (SOD), glutathione peroxidase (GPx) and catalase (CAT) activities, and reduced glutathione (GSH) content after  $\alpha$ -lipoic acid/ or  $\gamma$ -rays exposure in different rat groups.**

Groups	SOD	GPx	CAT	GSH
	$\mu\text{g/ g tissue}$	$\text{nmol/ g tissue}$	$\text{U/ g tissue}$	$\mu\text{mol/ g tissue}$
Control	$66.64 \pm 1.465$	$28.54 \pm 0.684$	$169.92 \pm 4.594$	$1.65 \pm 0.092$
$\alpha$ -lipoic acid	$64.21 \pm 1.577$	$29.54 \pm 0.738$	$172.21 \pm 4.471$	$1.63 \pm 0.044$
$\gamma$ -rays	$43.27 \pm 1.298^{a,b}$	$13.09 \pm 0.425^{a,b}$	$64.15 \pm 1.924^{a,b}$	$0.74 \pm 0.022^{a,b}$
$\alpha$ -lipoic acid+ $\gamma$ -rays	$62.45 \pm 1.686^c$	$24.64 \pm 0.641^c$	$130.92 \pm 3.272^{a,b,c}$	$1.42 \pm 0.035^{a,b,c}$

<sup>a</sup> significant difference when comparing with control group.

<sup>b</sup> significant difference when comparing with  $\alpha$ -lipoic acid-treated group.

<sup>c</sup> significant difference when comparing with  $\gamma$ -rays (6Gy) exposure group.



established antioxidant and anti-inflammatory properties,  $\alpha$ -lipoic acid is proposed to mitigate oxidative and inflammatory stress linked to cardiac dysfunction (Oskuye et al., 2024; Wang et al., 2025<sup>a</sup>). Given that  $\gamma$ -rays exposure induces oxidative stress and inflammation, this experimental model was utilised to simulate distinct cardiac injury (Karam et al., 2024).

Our findings revealed that  $\gamma$ -radiation exposure resulted in significant myocardial damage, as indicated by elevated levels of cardiac enzymes (LDH, CK-MB), pro-inflammatory markers (MPO, XO, MDA, PC, IL-6), and a notable decrease in endogenous antioxidants (SOD, GPx, CAT, FRA, GSH). However, co-administration of  $\alpha$ -lipoic acid significantly ameliorated these alterations, supporting its role as a potential radioprotective agent for cardiac tissues.

Oxidative stress and the accumulation of inflammatory markers significantly increase the risk of cardiac damage (Naderi et al., 2025).  $\gamma$ -rays exposure results in extensive injury to cardiac tissue, in line with previous studies (Ahmed et al., 2021; Karam et al., 2024; Thabet et al., 2022). In the present study,  $\alpha$ -lipoic acid conferred notable protection to cardiac tissues, mitigating the deleterious effects of  $\gamma$ -rays. This supports earlier findings that  $\alpha$ -lipoic acid reduces heart injury in rodent models (Nascimento et al., 2024; Oskuye et al., 2024).

FRA levels were significantly higher in the  $\alpha$ -lipoic acid group compared to the control. The beneficial effects of  $\alpha$ -lipoic acid may be attributed to its multifaceted ability to modulate antioxidant and pro-oxidant metabolism (Skibska et al., 2006). Due to its dithiol structure,  $\alpha$ -lipoic acid can scavenge various ROS, including hydroxyl radicals, superoxide anions, alkoxyl radicals, and peroxy radicals, thereby interrupting the propagation of lipid peroxidation (Biewenga et al., 1997). Beyond its direct scavenging activity,  $\alpha$ -lipoic acid participates in the regeneration of several endogenous antioxidants such as vitamin C, vitamin E, coenzyme Q10, and ubiquinone (Gotz et al., 1994; Kozlov et al., 1999; Moini et al., 2002). Additionally,  $\alpha$ -lipoic acid can indirectly elevate GSH levels by enhancing cysteine uptake, a rate-limiting step in GSH biosynthesis (Ruiter et al., 1981). This increase in thiol antioxidants contributes to an overall enhancement of total antioxidant capacity.

In this study, plasma FRA served as an

indicator of the oxidant/antioxidant balance. The total antioxidant capacity of plasma, assessed via FRA, was markedly reduced following  $\gamma$ -radiation-induced damage (Gupta et al., 2008; Verma et al., 2010). However,  $\alpha$ -lipoic acid administration effectively restored antioxidant status, protecting against lipid peroxidation in the red skeletal muscle of insulin-resistant rats (Dajnowicz-Brzezick et al., 2025).

The proinflammatory cytokine IL-6 adversely affects the structure of cardiomyocytes and contributes to heart muscle injury. Serum levels of IL-6 were measured to assess the acute inflammatory response following whole-body  $\gamma$ -rays exposure in rats (Moselhy et al., 2025).

The CK-MB is predominantly found in cardiac muscle, comprising up to 42% of total CK. Elevated CK-MB levels in the bloodstream are indicative of cardiac muscle injury (Zhang et al., 2025).

The current data demonstrate that oxidative stress significantly depletes SOD, leading to increased lipid peroxidation and elevated MDA concentrations. Similarly, other studies have shown that GSH levels may result from increased ROS production in cells and tissues (Li et al., 2025). In this study, early administration of  $\alpha$ -lipoic acid effectively counteracted  $\gamma$ -ray-induced lipid peroxidation and provided significant protection against endotoxin-induced oxidative stress in rats (Skibska et al., 2006).

Numerous studies have demonstrated the therapeutic potential of  $\alpha$ -lipoic acid in various disorders associated with oxidative stress (Hosny et al., 2024; Nascimento et al., 2024; Oskuye et al., 2024). Moreover,  $\alpha$ -lipoic acid has been shown to reduce IL-6 levels in several tissues, notably in cardiac tissue of rats (Khoder et al., 2022; Nemati et al., 2024). In the present study,  $\alpha$ -lipoic acid significantly restored CK-MB, MDA, IL-6, and SOD levels to values close to controls and effectively maintained GSH concentrations.

The observed elevation of LDH and CK-MB in irradiated rats indicates cardiac dysfunction, as these enzymes are released into circulation during heart injury. Treatment with  $\alpha$ -lipoic acid effectively normalized these enzyme levels (Hosny et al., 2024; Oskuye et al., 2024).

Increases in MPO and XO activities in damaged cardiac muscle corroborate previous findings (Hamed et al., 2024; Lu et al., 2025).

Administration of  $\alpha$ -lipoic acid mitigated  $\gamma$ -ray-induced oxidative injury by significantly enhancing antioxidant enzyme activities and restoring them near control levels (Petronilho *et al.*, 2016).

Exposure to  $\gamma$ -rays elevates ROS, resulting in increased  $H_2O_2$ , MDA, and PC levels (Rosen *et al.*, 2020).  $\alpha$ -Lipoic acid has shown protective effects against oxidative stress in patients with cardiac injuries and normalized PC levels close to baseline (Ratchford *et al.*, 2019; Wedan *et al.*, 2024). Furthermore, antioxidant enzymes GPx and CAT were reduced in irradiated rat hearts (Abdel-Magied *et al.*, 2020), whereas  $\alpha$ -lipoic acid treatment modulated these enzymes, alleviating  $\gamma$ -ray-induced damage (Kosoko *et al.*, 2017).

Overall,  $\alpha$ -lipoic acid may safeguard rat hearts from  $\gamma$ -ray-induced oxidative and inflammatory changes by preserving biomacromolecules and maintaining their native functional integrity under oxidative stress.

### **Conclusion**

The present study demonstrated that oral administration of  $\alpha$ -lipoic acid significantly mitigated the biochemical alterations induced by  $\gamma$ -radiation in rat cardiac tissue. Radiation exposure resulted in oxidative stress, inflammation, and elevated cardiac injury markers, while  $\alpha$ -lipoic acid supplementation effectively restored antioxidant enzyme activities and reduced oxidative and inflammatory biomarkers. These findings suggest that  $\alpha$ -lipoic acid offers a protective effect against radiation-induced cardiac injury, likely through its antioxidant and anti-inflammatory mechanisms. Therefore,  $\alpha$ -lipoic acid holds potential as a functional dietary supplement for reducing cardiac damage in contexts of radiation exposure. Further studies in humans are recommended to validate these results and explore clinical applicability.

### **Recommendation**

Further research needs to be carried out on animals and humans to provide additional understanding on the effects of  $\alpha$ -lipoic acid *in vivo*.

### **Limitations**

The study was conducted on rats, which may not fully replicate human physiological responses. Therefore, the findings may not be directly translatable to clinical applications in humans without further investigation. Only one dose of  $\gamma$ -radiation (6 Gy) was used. This dose

was chosen because it had previously shown biochemical effects on the heart of rats. The study does not address the effects of different doses or repeated exposure, which are often encountered in radiotherapy or environmental exposure. The study duration (10 days of treatment) limits the ability to assess long-term effects of  $\alpha$ -lipoic acid on cardiac tissue or its sustained protective action against radiation-induced damage. Although biochemical markers were measured, the study did not include detailed histological or morphological analysis of cardiac tissues to confirm tissue-level structural protection. The study used a fixed dose (100 mg/kg) without investigating dose-response relationships or evaluating the safety toxicity profile of varying doses. The focus was on oxidative stress and inflammation markers, without assessment of other relevant pathways like apoptosis, fibrosis, or endothelial dysfunction that may also play roles in radiation-induced cardiac injury. The study did not evaluate actual cardiac function (e.g., ECG, echocardiography) to support the biochemical findings with functional evidence. If only male rats were used, gender-specific responses to both radiation and  $\alpha$ -lipoic acid might have been overlooked.

Addressing these limitations in future studies would enhance the robustness and applicability of the findings.

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### **References**

- Abdel-Aziz N, Elkady AA, Elgazzar EM. (2021) Effect of Low-dose gamma radiation and lipoic acid on high- radiation-dose induced rat brain injuries. *Dose Response*. **19**(4):15593258211044845. doi: 10.1177/15593258211044845.
- Abdel-Magied N, Shedid SM. (2020) Impact of zinc oxide nanoparticles on thioredoxin-interacting protein and asymmetric dimethylarginine as biochemical indicators of cardiovascular disorders in gamma-irradiated rats. *Environ Toxicol*. **35**(4):430-442. doi: 10.1002/tox.22879.
- Adhab AH, Altalbawy FMA, Mahdi MS, Baldaniya

- L, Omar TM, Ganesan S, Juneja B, Pathak PK, Mansoor AS, Radi UK, Abd NS, Kadhim M. NADPH oxidases in cancer therapy-induced cardiotoxicity: mechanisms and therapeutic approaches. *Cardiovasc Toxicol*. 2025 **25**(4):631-649. doi: 10.1007/s12012-025-09976-4.
- Ahmed LA, Abdou FY, El Fiky AA, Shaaban EA, Ain-Shoka AA. Bradykinin-potentiating activity of a gamma-irradiated bioactive fraction isolated from scorpion (*Leiurus quinquestriatus*) venom in rats with doxorubicin-induced acute cardiotoxicity: Favorable modulation of oxidative stress and inflammatory, fibrogenic and apoptotic pathways. *Cardiovasc Toxicol*. 2021 **21**(2):127-141. doi: 10.1007/s12012-020-09602-5.
- Andreeva-Gateva P, Traikov L, Sabit Z, Bakalov D, Tafradjiiska-Hadjiolova R. Antioxidant effect of alpha-lipoic acid in 6-hydroxydopamine unilateral intrastratial injected rats. *Antioxidants* (Basel). 2020 **9**(2):122. doi: 10.3390/antiox9020122.
- Aziz MM, El-Sheikh MM, Mohamed MA, Abdelrahman SS, Mekawy MH. The senomorphic impact of astaxanthin on irradiated rat spleen: STING, TLR4 and mTOR contributed pathway. *Int J Immunopathol Pharmacol*. 2024 **38**:3946320241297342. doi: 10.1177/03946320241297342.
- Bergmeyer HU, Bergmeyer J, Grassl M. (1983) Methods of enzymatic analysis. 3, Weinheim: Verlag Chemie; .
- Biewenga GP, Haenen GR, Bast A. The pharmacology of the antioxidant lipoic acid. *Gen Pharmacol*. 1997 **29**(3):315-31. doi: 10.1016/s0306-3623(96)00474-0.
- Buege JA, Aust SD. (1978) Microsomal lipid peroxidation. *Methods Enzymol*. **52**:302-310. doi: 10.1016/s0076-6879(78)52032-6. PMID: 672633.
- Bustamante J, Lodge JK, Marcocci L, Tritschler HJ, Packer L, Rihn BH. (1998) Alpha-lipoic acid in liver metabolism and disease. *Free Radic Biol Med*. **24**(6):1023-39. doi: 10.1016/s0891-5849(97)00371-7.
- Cigala RM, Crea F, De Stefano C, Lando G, Milea D, Sammartano S. Modeling the acid-base properties of glutathione in different ionic media, with particular reference to natural waters and biological fluids. *Amino Acids*. 2012 **43**(2):629-48. doi: 10.1007/s00726-011-1110-0.
- Dajnowicz-Brzezic P, Żebrowska E, Maciejczyk M, Zalewska A, Chabowski A.  $\alpha$ -lipoic acid supplementation reduces oxidative stress and inflammation in red skeletal muscle of insulin-resistant rats. *Biochem Biophys Res Commun*. 2025 **742**:151107. doi: 10.1016/j.bbrc.2024.151107.
- Götz ME, Dirr A, Burger R, Janetzky B, Weinmüller M, Chan WW, Chen SC, Reichmann H, Rausch WD, Riederer P. Effect of lipoic acid on redox state of coenzyme Q in mice treated with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine and diethyldithiocarbamate. *Eur J Pharmacol*. 1994 **15**:266(3):291-300. doi: 10.1016/0922-4106(94)90139-2.
- Gupta ML, Sankhwar S, Verma S, Devi M, Samanta N, Agrawala PK, Kumar R, Singh PK. (2008) Whole body protection to lethally irradiated mice by oral administration of semipurified fraction of Podophyllum hexandrum and post irradiation treatment of Picrorhiza kurroa. *Tokai J Exp Clin Med*. 20:33(1):6-12. PMID: 21318957.
- Hamed MA, Adegboyega OO, Ojo OI, Akhigbe TM, Fajuyitan FD, (2024) Adeyemo OC, Odebunmi TF, Adeniyi OS, Omole IA, Akhigbe RE. Glutamine-mediated modulation of XO/uric acid/NF- $\kappa$ B signaling pathway ameliorates intestinal I/R-induced bacterial translocation and cardiorenal inflammatory injury. *Cell Biochem Biophys*. **82**(2):1007-1018. doi: 10.1007/s12013-024-01252-6.
- Helmy SA, Nour OA, G Abd El Salam AS. (2024) Ameliorative effect of Metformin / alpha-lipoic acid combination on diabetic nephropathy via modulation of YAP/ miR-29a/PTEN/p-AKT axis. *Int Immunopharmacol*. **135**:112294. doi: 10.1016/j.intimp.2024.112294.
- Hosny EN, Sawie HG, Abou-Seif HS, Khadrawy YA. Effect of caffeine-chitosan nanoparticles and  $\alpha$ -lipoic acid on the cardiovascular changes induced in rat model of obesity. *Int Immunopharmacol*. 2024 **129**:111627. doi: 10.1016/j.intimp.2024.111627.
- Hussien SM, Rashed ER. Immuno-biochemical impacts of gamma irradiation in male rats: a dose-response study. *Dose Response*. 2023 **21**(2):15593258231185461. doi: 10.1177/15593258231185461.
- Jermendy G, Rokszin G, Fábíán I, Kempler P, Wittmann I. (2023) Morbidity and mortality of patients with diabetic neuropathy treated with pathogenetically oriented alpha-lipoic acid versus symptomatic pharmacotherapies - A nationwide database analysis from Hungary. *Diabetes Res Clin Pract*.

- 201**:110734. doi: 10.1016/j.diabres.2023.110734.
- Jiang Q, Chen X, Gong K, Xu Z, Chen L, Zhang F. (2025) M6a demethylase FTO regulates the oxidative stress, mitochondrial biogenesis of cardiomyocytes and PGC-1 $\alpha$  stability in myocardial ischemia-reperfusion injury. *Redox Rep.* **30**(1):2454892. doi: 10.1080/13510002.2025.2454892.
- Kakkar P, Das B, Viswanathan PN. (1984) A modified spectrophotometric assay of superoxide dismutase. *Indian J Biochem Biophys.* **21**(2):130-132.
- Karam HM, Lotfy DM, A Ibrahim A, Mosallam FM, Abdelrahman SS, Abd-ElRaouf A. (2024) A new approach of nano-metformin as a protector against radiation-induced cardiac fibrosis and inflammation via CXCL1/TGF- $\beta$  pathway. *Naunyn Schmiedeberg's Arch Pharmacol.* **397**(9):6919-6927. doi: 10.1007/s00210-024-03052-4. Erratum in: *Naunyn Schmiedeberg's Arch Pharmacol.* 2024 397(9):7223. doi: 10.1007/s00210-024-03194-5.
- Khan M, Qiao F, Kumar P, Touhidul Islam SM, Singh AK, Won J, Singh I. (2022) Neuroprotective effects of Alda-1 mitigate spinal cord injury in mice: involvement of Alda-1-induced ALDH2 activation-mediated suppression of reactive aldehyde mechanisms. *Neural Regen Res.* **17**(1):185-193. doi: 10.4103/1673-5374.314312.
- Khoder NM, Sawie HG, Sharada HM, Hosny EN, Khadrawy YA, Abdulla MS. (2022) Metformin and alpha lipoic acid ameliorate hypothyroidism and its complications in adult male rats. *J. Diabetes Metab Disord.* **21**(2):1327-1337. doi: 10.1007/s40200-022-01063-7.
- Kosoko AM, Olurinde OJ, Akinloye OA. (2017) Doxorubicin induced neuro- and cardiotoxicities in experimental rats: Protection against oxidative damage by Theobroma cacao Stem bark. *Biochem Biophys Rep.* **10**:303-317. doi: 10.1016/j.bbrep.2017.01.012.
- Kozlov AV, Gille L, Staniek K, Nohl H. (1999) Dihydrolipoic acid maintains ubiquinone in the antioxidant active form by two-electron reduction of ubiquinone and one-electron reduction of ubisemiquinone. *Arch Biochem Biophys.* **363**(1):148-54. doi: 10.1006/abbi.1998.1064.
- Lawrence RA, Burk RF. (2012) Glutathione peroxidase activity in selenium-deficient rat liver. 1976. *Biochem Biophys Res Commun.* **425**(3):503-9. doi: 10.1016/j.bbrc.2012.08.016.
- Levine RL, Garland D, Oliver CN, Amici A, Climent I, Lenz AG, Ahn BW, Shaltiel S, Stadtman ER. (1990) Determination of carbonyl content in oxidatively modified proteins. *Methods Enzymol.* **186**:464-78. doi: 10.1016/0076-6879(90)86141-h.
- Li Y, Li M. (2025) Dihydromyricetin protects against hypoxia/reoxygenation injury in cardiomyocytes by activating miR-34a-mediated Notch1 pathway. *Cardiovasc Toxicol.* **25**(2):294-305. doi: 10.1007/s12012-025-09959-5.
- Liu C, Shi J, Xing L, Yao B, Liu J, Wang Y, Fan J. (2025) Discovery and confirmation of crucial genes associated with radiation-induced heart disease. *Int J Med Sci.* **22**(6):1278-1291. doi: 10.7150/ijms.107667.
- Lowry Oh, Rosebrough Nj, Farr Al, Randall RJ. (1951) Protein measurement with the Folin phenol reagent. *J Biol Chem.* **193**(1):265-275.
- Lu X, Liu F, Chen H, Cai H, Zhang L, Li J. (2025) Effects of WN1703 on Cardiovascular function in chronic hyperuricemia rats and myocardial injury mechanism exploration in H9C2 cells. *J Appl Toxicol.* **45**(3):418-431. doi: 10.1002/jat.4710.
- Mekkawy MH, Abdou FY, Ali MM, Abd-ElRaouf A. (2024) novel approach of using Maca root as a radioprotector in a rat testicular damage model focusing on GRP78/CHOP/Caspase-3 pathway. *Arch Biochem Biophys.* **755**:109963. doi: 10.1016/j.abb.2024.109963.
- Moini H, Packer L, Saris NE. (2002) Antioxidant and prooxidant activities of alpha-lipoic acid and dihydrolipoic acid. *Toxicol Appl Pharmacol.* **182**(1):84-90. doi: 10.1006/taap.2002.9437.
- Moselhy OA, Abdel-Aziz N, El-Bakhery A, Moselhy SS, Ibrahim EA. Curcumin nanoparticles alleviate brain mitochondrial dysfunction and cellular senescence in  $\gamma$ -irradiated rats. *Sci Rep.* 2025 15(1):3857. doi: 10.1038/s41598-025-87635-y.
- Naderi R, Seyhani A, Shirpoor A, Jafari A, Eyvani K. (2025) Effects of curcumin on cyclosporine A-induced oxidative stress, autophagy, and apoptosis in rat heart. *Mol Biol Rep.* **52**(1):310. doi: 10.1007/s11033-025-10334-4.
- Nascimento DVG, Alencar DF, da Silva MVB, Rocha DG, Roncari CF, Jorge RJB, Alves RS, David RB, Ferreira E Silva WT, Galindo LCM, de Queiroz TM. (2024) Cardiovascular and renal effects induced by alpha-lipoic acid treatment in two-kidney-one-clip hypertensive rats. *Biomedicine.* **12**(8):1751. doi: 10.3390/biomedicine12081751.
- Egypt. J. Rad. Sci. Applic.* **38**, No.1 (2025)



- Nemati S, Zavvari-Oskuye Z, Bafadam S, Mokhtari B, Badalzadeh R, Vakili A. (2024) Impact of combined alpha-lipoic acid and mitoquinone supplementation on myocardial infarction in aged rats: Heart performance and molecular mechanisms. *Exp Gerontol.* **189**:112402. doi: 10.1016/j.exger.2024.112402.
- Oskuye ZZ, Mehri K, Mokhtari B, Bafadam S, Nemati S, Badalzadeh R. (2024) Cardioprotective effect of antioxidant combination therapy: A highlight on MitoQ plus alpha-lipoic acid beneficial impact on myocardial ischemia-reperfusion injury in aged rats. *Heliyon.* **10**(6):e28158. doi: 10.1016/j.heliyon.2024.e28158.
- Pang Z, Li Q, Liu K, Wu X, Xu H, Chen Z, Dai H. (2024) Efficacy of melanin-loaded lipoic acid-modified chitosan hydrogel in diabetic wound healing. *Carbohydr Polym.* **340**:122215. doi: 10.1016/j.carbpol.2024.122215.
- Petronilho F, Florentino D, Danielski LG, Vieira LC, Martins MM, Vieira A, Bonfante S, Goldim MP, Vuolo F. (2016) Alpha-lipoic acid attenuates oxidative damage in organs after sepsis. *Inflammation.* **39**(1):357-365. doi: 10.1007/s10753-015-0256-4.
- Pingali U, Kammila S, Mekala P, Yareeda S, Penugonda S. (2024) A Study to Evaluate the Effect of Alpha-Lipoic Acid on Neuropathic Symptoms in Diabetic Neuropathy Patients on Gabapentin or Pregabalin. *Cureus.* 2024 Sep 27;**16**(9):e70299. doi: 10.7759/cureus.70299. PMID: 39469366; PMCID: PMC11513224.
- Ratchford SM, Clifton HL, Gifford JR, LaSalle DT, Thurston TS, Bunsawat K, Alpenglow JK, Richardson RS, Wright JB, Ryan JJ, Wray DW. (2019) Impact of acute antioxidant administration on inflammation and vascular function in heart failure with preserved ejection fraction. *Am J Physiol Regul Integr Comp Physiol.* **317**(5):R607-R614. doi: 10.1152/ajpregu.00184.2019.
- Rosen E, Kryndushkin D, Aryal B, Gonzalez Y, Chehab L, Dickey J, Rao VA. (2020) Acute total body ionizing gamma radiation induces long-term adverse effects and immediate changes in cardiac protein oxidative carbonylation in the rat. *PLoS One.* **15**(6):e0233967. doi: 10.1371/journal.pone.0233967.
- Ruiter DJ, van der Meulen J, Brouwer A, Hummel MJ, Mauw BJ, van der Ploeg JC, Wisse E. (1981) Uptake by liver cells of endotoxin following its intravenous injection. *Lab Invest.* **45**(1):38-45. PMID: 7253563.
- Said RS, Mohamed HA, Kassem DH. (2020) Alpha-lipoic acid effectively attenuates ionizing radiation-mediated testicular dysfunction in rats: Crosstalk of NF- $\kappa$ B, TGF- $\beta$ , and PPAR- $\gamma$  pathways. *Toxicology.* **442**:152536. doi: 10.1016/j.tox.2020.152536.
- Seyedpour N, Motevaseli E, Taeb S, Nowrouzi A, Mirzaei F, Bahri M, Dehghan-Manshadi HR, Zhaleh M, Rashidi K, Azmoonfar R, Yahyapour R, Najafi M. (2024) Protective effects of alpha-lipoic acid, resveratrol, and apigenin against oxidative damages, histopathological changes, and mortality induced by lung irradiation in rats. *Curr Radiopharm.* **17**(1):99-110. doi: 10.2174/0118744710244357231018070313.
- Skibska B, Józefowicz-Okonkwo G, Goraca A. (2006) Protective effects of early administration of alpha-lipoic acid against lipopolysaccharide-induced plasma lipid peroxidation. *Pharmacol Rep.* **58**(3):399-404. PMID: 16845214.
- Sztolsztener K, Hodun K, Chabowski A. (2022)  $\alpha$ -lipoic acid ameliorates inflammation state and oxidative stress by reducing the content of bioactive lipid derivatives in the left ventricle of rats fed a high-fat diet. *Biochim Biophys Acta Mol Basis Dis.* **1868**(9):166440. doi: 10.1016/j.bbadis.2022.166440.
- Tawfik SS, Elkady AA, El Khouly WA. (2019) Crocin mitigates  $\gamma$ -rays-induced hepatic toxicity in rats. *Environ Sci Pollut Res Int.* **26**(15):15414-15419. doi: 10.1007/s11356-019-04724-y.
- Thabet NM, Abdel-Rafei MK, Moustafa EM. (2022) Boswellic acid protects against Bisphenol-A and gamma radiation induced hepatic steatosis and cardiac remodelling in rats: role of hepatic PPAR- $\alpha$ /P38 and cardiac Calcineurin-A/NFATc1/P38 pathways. *Arch Physiol Biochem.* **128**(3):767-785. doi: 10.1080/13813455.2020.1727526.
- Titov VY, Osipov AN. (2017) Nitrite and nitroso compounds can serve as specific catalase inhibitors. *Redox Rep.* **22**(2):91-97. doi: 10.1080/13510002.2016.1168589.
- Tülüce Y, Osmanoğlu D, Rağbetli MÇ, Altındağ F. (2024) Protective action of curcumin and alpha-lipoic acid, against experimental ultraviolet-A/B induced dermal-injury in rats. *Cell Biochem Biophys.* **82**(4):3535-3546. doi: 10.1007/s12013-024-01442-2.

- Verma S, Gupta ML, Dutta A, Sankhwar S, Shukla SK, Flora SJ. Modulation of ionizing radiation induced oxidative imbalance by semi-fractionated extract of Piper betle: an in vitro and in vivo assessment. *Oxid Med Cell Longev*. 2010 3(1):44-52. doi: 10.4161/oxim.3.1.10349.
- Wang F, Li J, Zhang Z, Huang G, Zhang X, Liu Q, Xiao W, Liu F, Sun J, Liu Y, Ma Y, Zhuang R, Du Y, Wang X, Gao C, Gu X. Baicalin reduced vandetanib induced myocardial injury by regulating redox balance and NLRP3 inflammasome pathway. *Tissue Cell*. 2025<sup>a</sup> 94:102795. doi: 10.1016/j.tice.2025.102795.
- Wang X, Song SM, Lu WQ, Zhao Y, Lv RJ, He Y, Dong N, Yu Q, Yue HM. Alpha-lipoic acid alleviated intermittent hypoxia-induced myocardial injury in mice by promoting autophagy through Nrf2 signaling pathway. *Eur J Pharmacol*. 2025<sup>b</sup> 994:177380. doi: 10.1016/j.ejphar.2025.177380.
- Wang Y, Zheng Y, Qi B, Liu Y, Cheng X, Feng J, Gao W, Li T.  $\alpha$ -Lipoic acid alleviates myocardial injury and induces M2b macrophage polarization after myocardial infarction via HMGB1/NF- $\kappa$ B signaling pathway. *Int Immunopharmacol*. 2023 121:110435. doi: 10.1016/j.intimp.2023.110435.
- Wedan RJ, Longenecker JZ, Nowinski SM. Mitochondrial fatty acid synthesis is an emergent central regulator of mammalian oxidative metabolism. *Cell Metab*. 2024 36(1):36-47. doi: 10.1016/j.cmet.2023.11.017.
- Xie DM, Zhong Q, Xu X, Li Y, Chen S, Li M, Peng C. Alpha lipoic acid-loaded electrospun fibrous patch films protect heart in acute myocardial infarction mice by inhibiting oxidative stress. *Int J Pharm*. 2023 632:122581. doi: 10.1016/j.ijpharm.2023.122581.
- Xu J, Alameri AA, Zabibah RS, Gabr GA, Ramírez-Coronel AA, Bagheri H, Abedi-Firouzjah R. Protective potentials of alpha-lipoic acid against ionizing radiation-induced brain damage in rats. *Oxid Med Cell Longev*. 2023 2023:4999306. doi: 10.1155/2023/4999306.
- Zhang H, Dong J, Zhang J, Chen H, Liu T, Gan R, Wen J, Li Y. Effects of borneol on apoptosis of hypoxia/reoxygenation H9c2 cells and myocardial ischemia-reperfusion injury rats. *Acta Cir Bras*. 2025 40:e402225. doi: 10.1590/acb402225.
- Zhao Y, Guo M, Pei T, Shang C, Chen Y, Zhao L, Lu Y, Liang C, Wang J, Zhang J.  $\alpha$ -lipoic acid ameliorates arsenic-induced lipid disorders by promoting peroxisomal  $\beta$ -oxidation and reducing lipophagy in chicken hepatocyte. *Adv Sci (Weinh)*. 2025 12(11):e2413255. doi: 10.1002/advs.202413255.

## حمض ألفا ليبويك يحمي عضلة قلب الجرذ من الإصابة الناجمة عن أشعة جاما

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 القاهرة ، و2 قسم الوقاية من الإشعاع ، مركز بحوث الأمان النووي والأشعاعي ، ص.ب. ٧٥٥١ مدينة نصر  
 ، هيئة الطاقة الذرية المصرية ، القاهرة ، جمهورية مصر العربية.

يتواجد حمض ألفا ليبويك في أنسجة الثدييات والنباتات، ويتميز بتأثيرات مضادة للأكسدة والالتهابات. هدفت هذه الدراسة إلى تقييم قدرة حمض ألفا ليبويك، الذي تجرعه الجرذان عن طريق الفم، على الحماية من إصابة عضلة القلب الناجمة عن التعرض لجرعة ٦ جراي من أشعة جاما.

تم تعريض مجموعة من الجرذان لجلسة واحدة من إشعة جاما بجرعة 6 جراي، وتم مقارنة نتائج هذه المجموعة بنتائج المجموعة الضابطة، والمجموعة المعالجة بحمض ألفا ليبويك (جرعة ١٠٠ مجم/ كجم من وزن الجرذان لمدة ١٠ أيام) عن طريق أنبوب الفم، والمجموعة التي تعرضت لأشعة جاما وتم تجريعها بالحمض يوميًا لمدة ١٠ أيام.

أدى التعرض لأشعة جاما إلى زيادة في مستويات لاكتات ديهيدروجينيز (LDH) ، وميلوبيروكسيداز (MPO)، وأوكسيد الزانثين (XO) ، ومالونداي ألدهيد (MDA) ، وبروتين الكربونيل (PC) ، وكرياتين فوسفوكيناز - MB (CK-MB) ،

والإنترلوكين - ٦ (IL-6) ، وانخفاض ملحوظ في نشاط سوبر أوكسيد ديسميوتاز (SOD) ، والجلوتاثيون بيروكسيداز (GPx) ، والكاتالاز (CAT) ، ومقياس القدرة المضادة للأكسدة الكلية (FRAP) ، والجلوتاثيون المختزل (GSH) .

أدى تجريع الجرذان بحمض ألفا ليبويك بعد التعرض لأشعة جاما إلى تحسن في جميع المؤشرات المقاسة في أنسجة القلب والدم، مما يشير إلى حماية واستعادة لأنظمة الدفاع البيولوجية المضادة للأكسدة.

الخلاصة: يمكن استخدام حمض ألفا ليبويك كمكمل غذائي ووقائي للتخفيف من إصابة عضلة قلب الجرذ الناجمة عن التعرض لأشعة جاما، ويعزى تأثيره إلى خصائصه المضادة للأكسدة والالتهابات.